SunLong Biotech Co.,LTD

Tel: 0086-571- 56623320 Fax:0086-571- 56623318

E-mail:sales@sunlongbiotech.com

www.sunlongbiotech.com

Quick Step Camel Theileria IgM,Thi-IgM ELISA Kit

Size: 96 T, 48T

Catalogue Number:QS0061Cm

Assay Time: 60 minutes

Store all reagents at 2-8℃/-20°C

Validity Period: 2-8℃ for six months, -20°C for one year. Avoid repeated thaw cycles.

For samples: In serum, plasma, culture media or any biological fluid.

FOR RESEARCH USE ONLY！

NOT FOR THERAPEUTIC OR DIAGNOSTIC APPLICATIONS！

PLEASE READ THROUGH ENTIRE PROCEDURE BEFORE BEGINNING！

Quick Step Camel Theileria IgM,Thi-IgM ELISA Kit

FOR RESEARCH USE ONLY

Purpose

Our Quick Step Camel Theileria IgM,Thi-IgM ELISA Kit is to for the qualitative determination of Thi-IgM in Camel serum, plasma, culture media or any biological fluid.

Principle

This ELISA kit uses Sandwich-ELISA as the method. The Microelisa stripplate provided in this kit has been pre-coated with an antigen specific to Thi-IgM. Standards or samples are added to the appropriate Microelisa stripplate wells and combined to the specific antigen. Then a Horseradish Peroxidase (HRP)-conjugated antigen specific for Thi-IgM is added to each Microelisa stripplate well and incubated. Free components are washed away. The TMB substrate solution is added to each well. Only those wells that contain Thi-IgM and HRP conjugated Thi antigen will appear blue in color and then turn yellow after the addition of the stop solution. The optical density (OD) is measured spectrophotometrically at a wavelength of 450 nm. The presence of Thi-IgM is determined by comparing with the CUT OFF value.

Materials provided with the kit

	Materials provided with the kit	96 determinations	48 determinations
1	User manual	1	1
2	Closure plate membrane	2	2
3	Sealed bags	1	1
4	Microelisa stripplate	1	1
5	Negative control	0.5ml×1 bottle	0.5ml×1 bottle
6	Positive control	0.5ml×1 bottle	0.5ml×1 bottle
7	HRP-Conjugate reagent	6ml×1 bottle	3ml×1 bottle
8	Sample diluent	6ml×1 bottle	3ml×1 bottle
9	Chromogen Solution A	6ml×1 bottle	3ml×1 bottle

10	Chromogen Solution B	6ml×1 bottle	3ml×1 bottle
11	Stop Solution	6ml×1 bottle	3ml×1 bottle
12	Wash Solution	20ml (30X)×1bottle	20ml (20X)×1bottle

Sample collection and storages

1. Can't detect the samples which contain NaN3, because NaN3 inhibits HRP activity of the horseradish peroxidase.
2. Extract as soon as possible after Specimen collection, Extracted according to the relevant literature.

Cell culture supernates and plant exact fluids - Remove particulates by centrifugation and assay immediately or aliquot and store samples at -20°C or -80°C. Avoid repeated freeze-thaw.

Notes:

Sample extraction and ELISA assay should be performed as soon as possible after sample collection. The samples should be extracted according to the relevant literature. If ELISA assay can not be performed immediately, samples can be stored at -20 ℃ . Repeated freeze-thaw cycles should be avoided.

Procedure

1. In the Microplate, number the corresponding micropores of the sample in sequence, leave two wells as negative control, two wells as positive control and one empty well as blank control. (Do not add samples and HRP-Conjugate reagent into blank control well, same for the rest steps)
2. **Add Samples:**Add 50µl of negative control and positive control into the designated negative control well and positive control well, respectively. Add 40µl Sample diluent and 10µl sample into the sample wells. Samples should be loaded onto the bottom without touching the well wall. Mix well with gentle shaking.

3. **Add HRP:**Add 50μl HRP-Conjugate reagent to each well except the blank control well.
4. **Incubation:** Incubate 30 min at 37℃ after sealed with Closure plate membrane.
5. **Preparing solution:**Dilute the concentrated Washing Solution with distilled water (30 times for 96T and 20 times for 48T).
6. **Washing:**Carefully peel off Closure plate membrane, aspirate and refill with Wash Solution (350μl to 400μl, or fill it completely, overflow is acceptable). Remove the Wash Solution after resting for 30 seconds. Repeat the washing procedure for 5 times. After the last wash, remove any remaining Wash Solution by aspirating or decanting. Invert the plate and blot it against clean paper towels.
7. **Chromogenic reaction:**Add 50μl Chromogen Solution A and 50μl Chromogen Solution B to each well, mix with gently shaking and incubate at 37℃ for 10min. **Protect from light.**
8. **Stop the reaction:**Add 50μl stop solution to each well to terminate the reaction. The color in the well should change from blue to yellow.
9. **Assay:**Read absorbance O.D. at 450nm using a Microtiter Plate Reader. The OD value of the blank control well is set as zero. Assay should be carried out within 15 minutes after adding stop solution.

Notes:

1. Store the kit at 4 ℃ upon receipt. The kit should be equilibrated to room temperature before the assay. Remove any unneeded strips from Thi antigen-Coated plate (used for detecting Thi-IgM), reseal them in zip-lock foil and keep at 4 ℃.
2. Precipitates may appear in concentrated washing buffer. Please heat the buffer to dissolve all the precipitates, which will not affect the results.
3. In order to avoid cross-contamination, Closure plate membranes are for one-time use only.
4. Please keep Substrate away from light.
5. All the operation should be accordance with the manufacturer's instructions strictly. The

results determined by the Microelisa stripplate Reader.

6. All the samples, washing buffer and wastes should be treated as infectious agents.
7. Reagents from different lots should not be mixed.

Determine the result

Test effectiveness: the average value of positive control ≥1.00; The average value of negative control ≤0.10.

The critical value (CUT OFF) calculation: critical value = the average value of negative control + 0.15

Negative judgement: if the OD value<CUT OFF, the sample is Camel Thi-IgM negative.

Positive judgement: if the OD value ≥CUT OFF, the sample is Camel Thi-IgM positive.

Storage and validity

1. Storage: 2-8℃/-20°C.
2. Duration: 2-8℃ for six months, -20°C for one year. Avoid repeated thaw cycles.